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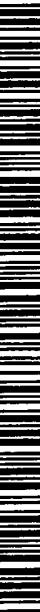
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(54) Title: SOLUTION FOR TREATING AUTOLOGOUS TISSUE FOR IMPLANTATION

(57) Abstract: An aqueous solution containing a water miscible organic solvent, polyethylene glycol and heparine, is used to modify the tissue reactivity of an autologous tissue freshly obtained from a host mammal and to render the tissue temporarily more rigid than in its native state, and better suited for shaping, moulding, handling and cutting prior to implantation into the host patient. The implant is highly resistant or immune to thickening, contraction and reduced fibrin deposition after it is implanted and exposed to the bloodstream of the host.

1       **SOLUTION AND METHOD FOR TREATING AUTOLOGOUS**  
2                   **TISSUE FOR IMPLANT OPERATION**  
3                   **BACKGROUND OF THE INVENTION**

4       **1. Field of the Invention**

5               The present invention is in the field of methodology and materials for  
6               performing autologous tissue transplants. More particularly, the present  
7               invention is directed to a solution suitable for treating autologous tissue after  
8               the tissue has been removed from a mammal, to render the tissue more suitable  
9               for handling and molding it into the desired shape and obtain predictable  
10               results after implantation in the mammal.

11       **2. Brief Description of the Prior Art**

12               Significant advances have been made in the field of treatment of  
13               defective heart valves due to abnormalities during fetal development, or due to  
14               infectious or degenerative diseases. These surgical treatments most often  
15               require the use of biocompatible materials that can be either synthetic  
16               polymers or of biological origin, either from the patient (autologous), an  
17               individual of the same species (homologous), or different species  
18               (heterologous or xenograft). Defective heart valves are replaced with  
19               mechanical valves or tissue valves, such as cadaver or animal aortic valves  
20               (bioprostheses). Because of their more-or-less predictable mechanical wear  
21               properties, the mechanical prostheses have been proven suitable for their  
22               intended purpose, primarily in younger patients. However, mechanical  
23               prostheses have their disadvantages because patients having them require  
24               long-term, constant and vigorous anti-coagulant therapy. In older patients  
25               however the bioprostheses have been favored mainly because they do not  
26               require anti-coagulation therapy, and in older patients they do not tend to  
27               undergo calcification as often as they tend to do in younger patients. Cryo-  
28               preserved homografts have been used widely in the western world during the  
29               recent years. However, these are hard and expensive to obtain, ship and store,

1 and their availability on a world-wide basis appears to be limited.

2 As is known, heart valve repair or replacement and many other implant  
3 operations require soft connective tissue which in preparation for implantation  
4 needs to be sized and cut into specific shapes. A substitute for such soft  
5 connective tissue of biological origin can be provided by flat sheets of certain  
6 synthetic materials. However, it is difficult to find synthetic materials which  
7 can match the compliance of the native tissue they are intended to replace, and  
8 which do not engender adverse reaction by the recipient of the implant.

9 Autologous tissues, such as pericardium, hold the promise for an ideal soft  
10 tissue replacement material in implants, and fresh autologous pericardium has  
11 been used in the prior art as a tissue source for repairing a variety of heart  
12 lesions, including heart valves. However, results with the use of such fresh  
13 untreated autologous tissue were less than satisfactory because tissue  
14 contraction distorted the repair, and in case of heart valves, tended to render  
15 the leaflets non-functional some time after operation. Generally speaking, the  
16 problem with fresh autologous soft connective tissue, such as pericardium, is  
17 that such tissues are often too soft and flexible to cut and otherwise handle  
18 especially during open heart surgery where an atmosphere of urgency prevails.

19 As an improvement Dr. Duran (one of the inventors of the present invention)  
20 developed a procedure in which the autologous tissue that has been freshly  
21 obtained from the patient operated on, is treated with 0.5% glutaraldehyde in a  
22 mold for 10 minutes. Thereafter, it is cut into the desired shape dictated by  
23 the mold and is placed in the patient as new replacement heart valve leaflets.  
24 Although this procedure works reasonably well, the disadvantage of tissues  
25 treated by glutaraldehyde is that, similarly to xenografts, such tissues may  
26 well undergo calcification in long term implants.

27 The present invention provides an alternative to glutaraldehyde fixation  
28 of autologous tissues and yet eliminates the problems caused by contraction of  
29 fresh tissue and the difficulty of handling and manipulating soft tissue.

1 Because the invention avoids the above-noted problems by treating the  
2 autologous tissue with an aqueous solution of alcohols and other materials,  
3 prior art describing solutions and methods for treating biological tissues and  
4 specimens are thought to be of interest as background to the present invention.  
5 Such prior art can be found in United States Patent Nos. 5,558,875;  
6 5,296,514; 5,276,006; 4,323,358 and 4,329,492. Among the foregoing, the  
7 most recently issued United States Patent No. 5,558,875 describes a process  
8 of preparing a collagenous prosthesis by soaking tissue in an organic detergent  
9 for sufficient time to disrupt the cell membrane and to solubilize the cellular  
10 membrane proteins of the collagenous tissue and thereafter extracting and  
11 removing the cellular membrane proteins from the collagenous tissue by  
12 mechanical washing to obtain the prosthesis and thereafter preserving the  
13 prosthesis in alcohol. The process is said to preserve the elasticity of the  
14 prosthesis.

15 The following articles or scientific publications also provide  
16 background of interest to the present invention: *Chachques et al.*, Ann. NY  
17 Acad Sci. 1988, 529:184; *Love et al.*, J. Heart Valve Dis 1992: 1:232-41;  
18 *Chauvaud et al.*, J. Thorac Cardiovasc Surg. 1991, 102:171-8; *Duran et al.*, J.  
19 Thorac Cardiovasc Surg. 1995, 11-511-6; *Vyawahare et al.*, 4th Scientific  
20 Meeting International Association for Cardiac Biological Implants,  
21 Washington DC, May, 1997; *Ritter et al.*, Plastic & Reconstructive Surgery.  
22 101(1):142-6, Jan., 1998; and *Vetter et al.*, J. Thorac Cardiovasc Surg,  
23 35(1):11-5, Feb. 1987.

## 24 SUMMARY OF THE INVENTION

25 It is an object of the present invention to provide a composition  
26 (solution) and method for treating autologous soft tissue so as to render it  
27 easier to handle and shape for implantation, while avoiding disadvantages  
28 caused by aldehyde treatment of such tissue.

29 It is another object of the present invention to provide a composition

1 (solution) and method that meets the foregoing objective and which treats the  
2 autologous soft tissue during a surgical procedure and while said procedure is  
3 in progress.

4 The foregoing and other objects and advantages are attained by  
5 exposing for approximately 2 to 8 minutes a fresh autologous tissue, such as  
6 pericardium, to an aqueous solution containing approximately 10 to 70 % by  
7 volume of a water-miscible non toxic polar solvent, such as ethyl alcohol,  
8 approximately 2 to 30 % by weight of polyethylene glycol of a molecular  
9 weight between approximately 6,000 to 15,000 D, and approximately 0.01 to  
10 1.0 % by weight of heparin. The tissue preferably, and most frequently in  
11 accordance with the procedure is immersed in the above-described solution  
12 while placed in a suitable mold. In case of preparing the tissue for heart valve  
13 replacement the mold is configured to provide the appropriate shape and  
14 dimension for the replacement heart valve leaflets. The soft tissue implant  
15 treated in the foregoing manner temporarily becomes more rigid and easier to  
16 handle during surgical procedure than unprepared fresh tissue. However,  
17 within approximately the time taken to perform the surgical procedure of  
18 implantation the treated tissue regains its original physical properties,  
19 including its elasticity.

20 **BRIEF DESCRIPTION OF THE DRAWING FIGURES**

21 **Figure 1** is a schematic perspective view showing the general  
22 configuration of one cusp of a negative template of a mold used for shaping  
23 an aortic valve replacement from autologous tissue, utilizing the novel solution  
24 and method of the present invention.

25 **Figure 2** is a schematic perspective view showing the general  
26 configuration of one cusp of a positive template of the mold used for shaping  
27 an aortic valve replacement from autologous tissue, utilizing the novel solution  
28 and method of the present invention.

29 **Figure 3** is a top plan view of the negative cusp of **Figure 1**.

1       **Figure 4** is an end plan view of the negative cusp of **Figure 1**.

2       **Figure 5** is a side plan view of the negative cusp of **Figure 1**.

3       **Figure 6** is a schematic top plan view showing the general  
4       configuration of three negative cusps of **Figure 1** assembled to form a  
5       negative template used for shaping an aortic valve replacement from  
6       autologous tissue, utilizing the novel solution and method of the present  
7       invention.

8       **Figure 7** is a front plan view of the negative template of **Figure 6**.

9       **Figure 8** is a schematic perspective view of the negative template of  
10      **Figure 6**.

11      **Figure 9** is a detailed top plan view of a first preferred embodiment of a  
12       negative template used for shaping a pericardial valve replacement from  
13       autologous tissue, utilizing the novel solution and method of the present  
14       invention.

15      **Figure 10** is an end view of the first preferred embodiment of the  
16       negative template of **Figure 9**.

17      **Figure 11** is a front plan view of the first preferred embodiment of the  
18       negative template of **Figure 9**, with part of the front material broken away.

19      **Figure 12** is a detailed top plan view of a first preferred embodiment of  
20       a positive template used for shaping a pericardial valve replacement from  
21       autologous tissue, utilizing the novel solution and method of the present  
22       invention.

23      **Figure 13** is an end view of the first preferred embodiment of the  
24       positive template of **Figure 12**.

25      **Figure 14** is a front view of the first preferred embodiment of the  
26       positive template of **Figure 12**.

27      **Figure 15** is a perspective view of the first preferred embodiment of the  
28       negative template of **Figure 9**.

29      **Figure 16** is a perspective view of the first preferred embodiment of the

1 positive template of **Figure 12**.

2 **Figure 17** is a top plan view showing the first preferred embodiment of  
3 the negative template of **Figure 9** and the first preferred embodiment of the  
4 positive template of **Figure 12** assembled to one another.

5 **Figure 18** is a front plan view showing the first preferred embodiment  
6 of the positive template of **Figure 9** and the first preferred embodiment of the  
7 negative template of **Figure 12** assembled to one another.

8 **Figure 19** is a partial cross-sectional view taken on lines 19,19 of  
9 **Figure 18**.

10 **Figure 20** is a view showing the assembled mold of **Figure 18** having  
11 autologous tissue and immersed in a solution in accordance with the present  
12 invention.

13 **Figure 21** is a partial cross sectional view of the mold with autologous  
14 tissue, the cross-section being taken on lines 21,21 of **Figure 20**.

15 **Figure 22** is a partial schematic view, schematically showing the  
16 trimming of excess autologous tissue to form a replacement heart valve, in  
17 accordance with the present invention.

18 **Figure 23** is a cross-sectional view taken on lines 23,23 of **Figure 22**.

19 **Figure 24** is a partial cross sectional view of the trimmed autologous  
20 tissue.

## 21 DESCRIPTION OF THE PREFERRED EMBODIMENTS

22 The present invention is practiced in conjunction with a surgical  
23 procedure wherein a heart valve, aortic, pulmonary, tricuspid or mitral, or  
24 other biological membrane is replaced or repaired. Because its most frequent  
25 use is in conjunction with replacement of defective membranes in the heart,  
26 the present invention is described here primarily as it pertains to replacement  
27 of defective heart valves. In accordance with the present invention the  
28 operating surgeon excises a membrane-like fresh autologous tissue from the  
29 patient and by application of the solution of the present invention changes the

1 physical properties of the fresh autologous tissue to be better suited for  
2 trimming, handling, and manipulation during implantation into the patient.  
3 Membrane-like tissues which are suitable to be handled and implanted in  
4 accordance with the present invention include the peritoneum, pericardium,  
5 gut, dermis pleura and tendon. For open heart surgery and replacement of  
6 defective heart valves the use of the patient's pericardium is preferred, and  
7 therefore the invention is described herein primarily in connection with the use  
8 of pericardium as the membrane-like autologous tissue.

9 In accordance with the invention, the freshly obtained pericardium (or  
10 other membrane-like autologous tissue) is treated with an aqueous solution  
11 containing approximately 10 to 70 % by volume of a water-miscible non-toxic  
12 organic solvent, approximately 2 to 30 % by weight of polyethylene glycol of  
13 a molecular weight between approximately 6,000 to 15,000 D, and  
14 approximately 0.01 to 1.0 % by weight of heparin, the rest of the solution  
15 being water. Examples of suitable water-miscible organic solvents or liquids  
16 are lower alkyl, especially C<sub>1</sub> to C<sub>3</sub> alcohols, such as methanol, ethanol and  
17 *iso*-propanol, and acetone, acetonitrile and methyl ethyl ketone. A more  
18 preferred range of the components in the solution in accordance with the  
19 present invention is approximately 15 to 60 % by volume of the water-  
20 miscible organic liquid, 2 to 10 % by weight of polyethylene glycol, and 0.1 to  
21 0.7 % by weight of heparin.

22 Preferably the organic solvent is ethyl alcohol, and in the presently  
23 most preferred embodiment of the solution there is approximately 50 % by  
24 volume ethanol, approximately 5 % by weight of polyethylene glycol having a  
25 molecular weight of approximately 8,000 D, and approximately 0.5 % by  
26 weight of heparin. The biological membrane is thoroughly exposed to the  
27 solution for sufficient time to provide the desired results of rendering the  
28 membrane more rigid and therefore easier to trim, suture and otherwise  
29 handle. However, usually a time limit is set to this exposure by the fact that

1 the process occurs while the patient is undergoing surgery, usually open heart  
2 surgery. It was found in practice that approximately 2 to 8 minutes of  
3 exposure of the biological membrane to the solution is sufficient.  
4 Nevertheless, under circumstances where the surgical procedure *per se* does  
5 not represent a time-limiting factor, the biological membrane can be kept in  
6 the solution for indefinite length of time provided the solution is kept under  
7 sterile condition. Treatment by this solution kills the living cells in the  
8 membrane although treatment with the solution containing organic solvent at  
9 the lower end of the above-described range may only kill cells on the surface  
10 of the membrane and merely retards the biological response of cells in the  
11 interior. Nevertheless, unlike treatment with glutaraldehyde, treatment with  
12 the solution of the present invention does not result in any cross-linking of the  
13 membrane materials. The biological membrane or tissue becomes more rigid  
14 or stiff during exposure to the solution partly because of the hypertonic,  
15 dehydrating nature of the solution.

16 Hardening or stiffening of the membranes is temporary, however,  
17 because after sufficient rinsing with saline or upon equilibration with isotonic  
18 biological fluids, such as blood, the biological membranes regain virtually  
19 completely their original physical properties, and as a result are well suited for  
20 their intended function as replacement of natural membranes, primarily as  
21 heart valves.

22 A preferred manner of practicing the present invention, together with  
23 molds that are used for shaping pericardium or other biological membranes to  
24 provide aortic and pericardial heart valve replacements are illustrated in the  
25 drawing figures. Referring now back to the Brief Description of the Drawing  
26 Figures, **Figures 1 through 8** schematically illustrate the basic geometry of a  
27 mold comprising a negative 30 and a positive 32 template for forming an  
28 aortic valve replacement in accordance with the present invention. **Figures 1**  
29 through 5 schematically illustrate the basic geometry of the individual negative

1    34 and positive 36 cusps of the templates 30 and 32 that together form the  
2    mold. The templates 30 and 32 are made from thin plastic material, and are  
3    configured and dimensioned to provide the aortic heart valve replacement for  
4    the individual patient who is being operated on. As it will be readily  
5    understood by those skilled in the art of cardiology and related cardiac  
6    surgery, primarily echocardiograms of the patient provide the information as  
7    to what size heart valve replacement is needed. Edges of the templates 30 and  
8    32 are beveled or rounded in order to facilitate trimming of excess tissue with  
9    a surgical knife.

10       In accordance with one manner of practicing the invention, the  
11    pericardium (or other suitable biological membrane) is placed into the mold  
12    between the negative and positive templates and treated for approximately 5  
13    minutes with the solution of the invention by immersion in the solution.  
14    Thereafter it is very quickly (for less than 5 seconds) rinsed with saline  
15    solution containing approximately 250 unit per ml heparin. This step of  
16    treating the tissue with heparin solution for a very brief period of time is not  
17    necessary for the successful practice of the invention and is therefore optional.  
18    In any event, after removal from the mold and having been treated with the  
19    solution of the invention the tissue is more rigid than the native untreated  
20    pericardium, and is easier to handle. Excess tissue is then removed by  
21    trimming with a surgical knife, the tissue in the shape of heart valve leaflets is  
22    removed from the mold, and is thereafter surgically implanted. The  
23    increased rigidity or stiffness of these replacement leaflets renders the  
24    implantation procedure easier to handle. After suturing is completed, the  
25    tissue is irrigated with saline solution, whereupon it regains its original  
26    physical properties. As noted above, the biological membrane treated in  
27    accordance with the present invention regains its original physical properties  
28    upon adequate rinsing with saline, or achieving equilibrium with isotonic  
29    aqueous fluid, such as blood.

1       **Figures 9 through 24 illustrate in more detail an actual mold**  
2       comprising a negative template 38 and a positive template 40 adapted for  
3       shaping a flat biological membrane, such as pericardium, to form replacement  
4       leaflets for a pericardial valve. The two templates 38 and 40 of this mold are  
5       made of thin plastic material having beveled edges, and the templates are  
6       dimensioned to provide replacement leaflets of appropriate size for the patient  
7       who is undergoing the open heart surgery. For use in conjunction with the  
8       present invention, the negative 38 and the positive template 40 both are  
9       provided with a plurality of apertures 42. When this mold is used in the  
10      practice of the present invention, the autologous biological membrane,  
11      preferably pericardium excised from the patient who is undergoing open heart  
12      surgery, is placed between the templates, 38 and 40, that is into the mold.  
13      Accordingly, the pericardium, shown in **Figures 21 through 24 as 44** is  
14      sandwiched between the two templates 38 and 40. The two templates of the  
15      mold are held together with suitable plastic clips 46, shown in **Figure 20**.  
16      The mold including the pericardium 44 is then immersed for approximately 5  
17      minutes in the solution 48 of the invention, as is schematically shown in  
18      **Figure 20**. During this time the solution 48 percolates through the apertures  
19      42 into the pericardium 44 and renders the pericardium 44 more rigid than in  
20      its natural native state. After removal from the solution, the tissue 44 is  
21      trimmed with a surgical instrument along the beveled edges of the mold. The  
22      surgical instrument is schematically shown in **Figure 22** and bears the  
23      reference numeral 50. The resulting replacement heart leaflets (not shown) are  
24      then removed from the mold, and may be quickly (less than 5 seconds) rinsed  
25      with saline solution containing approximately 250 unit per ml heparin. This  
26      optional quick rinsing does not yet decrease the rigidity of the tissue which  
27      was the result of treatment with the solution. Surgical implantation of the  
28      replacement leaflets is greatly facilitated by its increased rigidity or stiffness.  
29      After implantation, the replacement leaflets are irrigated with saline and regain

1 their original physical properties. A substantial advantage of the heart valve  
2 replacements obtained in accordance with the present invention is that, unlike  
3 replacement valves made of autologous tissues in the prior art, the replacement  
4 valves of the invention do not contract or shrink after implantation. Generally  
5 speaking, implants of autologous tissues which have been treated in  
6 accordance with the present invention are highly resistant or immune to  
7 thickening, contraction or fibrin deposition after the implants are placed into  
8 the bloodstream of the host. The above described process of exposing the  
9 biological membrane to the solution of the invention while the membrane is  
10 held in an appropriately configured and dimensioned mold is the presently  
11 preferred mode of practicing the invention.

12 **Specific Examples**

13 Fresh autologous tissues were dissected from the sheep and placed on  
14 molds which were used as templates for cutting the tissues into a shape which  
15 appeared as three half-moons joined together at the base of the half-moon.  
16 This particular shape was designed for the purpose of aortic or pulmonary  
17 heart valve cusp extension operation. Pericardial tissues cut according to this  
18 design can be directly implanted into the heart of the individual patient where  
19 the tissue is derived from. The tissues in the mold were immersed into a  
20 solution containing 50 % by volume of alcohol, 5 % by weight of polyethylene  
21 glycol (MW=8,000) and 0.5 % by weight of heparin for five minutes at room  
22 temperature. The tissues in the mold were removed from the solution and  
23 after the removal of excess liquid outside the tissues, the tissues were rinsed in  
24 saline containing 250 unit/ml of heparin for less than 5 seconds.

25 Tissues treated with the solution mentioned above appeared to be  
26 slightly translucent and natural in color. Unlike the fresh untreated tissues, the  
27 treated tissues were stiffer so that it was relatively easy to lift the tissues  
28 without the tissues folding onto themselves. The treated tissues were easily  
29 spread on a flat or curved surface with different markings for cutting the

1 tissues. Since the cut tissues maintained their shape, they were easily  
2 implanted as replacement leaflets of heart valves. Yet, once the suturing of the  
3 tissues was completed and a small amount of saline was irrigated onto the  
4 implanted tissues, the tissues became soft quickly. Within the time required to  
5 close the open heart, the physical properties of the tissues became  
6 indistinguishable from the fresh untreated tissues. Therefore the resulting  
7 implants function perfectly as repaired heart valves.

8 Human fibroblasts and umbilical cord vein endothelial cells were  
9 cultured on the treated tissues after they were rinsed in saline containing 250  
10 unit/ml of heparin to study their biocompatibility. Round discs of the tissues  
11 were cut to fit the bottom of the wells of a 24 well culture plate. Tygon<sup>R</sup>  
12 flexible rings were placed on top of the tissues to ensure a good seal at the  
13 edge of the tissues. Cells were seeded on the tissues in normal culture media  
14 for one week. At the end of the incubation period, tissues were recovered and  
15 processed for histology. Both human umbilical cord vein endothelial cells and  
16 human skin fibroblasts attached and proliferated on the treated tissues as  
17 evidence that after rinsing in saline the treated tissue is not cytotoxic and  
18 biocompatible for host cells to attach and proliferate. The attachment and  
19 proliferation of endothelial cells and other connective tissue cells on cardiac  
20 implants is potentially important for the long term survival of the implant.

21 Integrity of the collagen fibers in the treated tissues was examined by  
22 melting temperature measurements. Tissues were heated in phosphate  
23 buffered saline from 37° C until they shrunk. The shrinkage temperature of  
24 the treated tissues after they were rinsed in saline was 64±1° C which is  
25 identical to untreated fresh tissue indicating that the collagen fibers remained  
26 intact throughout the treatment and saline-rinse process.

27 Efficacy of the treated tissues as useful cardiovascular implants was  
28 tested by implanting the treated autologous tissues in the descending aorta of  
29 sheep in different configurations. A piece of pericardium was dissected and

1 divided into three pieces with different shapes, namely a trapezoid, a strip  
2 made into a conduit and a square. These pieces of tissues were implanted  
3 serially in the descending aorta of sheep. The trapezoid shaped tissue was  
4 implanted upstream as a patch on the aortic wall, next to it down-stream the  
5 short conduit was implanted and further down-stream a square shaped tissue  
6 was placed as a semi-free flap across the lumen inside the aorta with two edges  
7 of the square attached to opposite side of the inner wall of the aorta. When  
8 fresh autologous tissues were implanted under the same condition, the semi-  
9 free flap in the aorta lumen became fibrotic and contracted within 30 days.  
10 However the patch and the conduit upstream, that was implanted in  
11 accordance with the present invention did not show the same reaction. There  
12 was also evidence of thrombus formation and fibrin deposition on the surfaces  
13 of the fresh implants. When the treated implants (not rinsed in saline) were  
14 implanted in sheep in the same manner all implants remained intact after 30  
15 days without any evidence of fibrotic reaction and tissue contraction.  
16 Thrombus and fibrin deposition were minimal or absent on these implants.

17 In still other further examples fresh bovine pericardial tissues were  
18 treated using the solution of the invention. The treated tissues were cut and  
19 trimmed to sizes and shapes suitable for valve repairs. The treated and  
20 trimmed tissues were sutured in the aortic roots of isolated human and porcine  
21 hearts. The whole hearts were then mounted on a pulse duplicator to examine  
22 the competency of the repair valve. The treated tissues were very flexible and  
23 the reconstructed valves functioned normally as competent aortic valves.

## WHAT IS CLAIMED IS:

2 1. A liquid composition adapted for treating autologous biological  
3 tissue for modifying its tissue reactivity and for rendering the tissue  
4 temporarily more rigid than in its natural state, the composition comprising:

5 approximately 10 to 70 % by volume of a water miscible non-toxic  
6 organic solvent selected from the group consisting of an alcohol having 1 to 3  
7 carbons, acetone, acetonitrile and methyl ethyl ketone;

8 approximately 2 to 30 % by weight of polyethylene glycol having a  
9 molecular weight in the range of approximately 6,000 to 15,000 D;

10 approximately 0.01 to 1.0 % by weight of heparin, and

11 the balance of the composition substantially consisting of water.

12        2.      The liquid composition of Claim 1 wherein the water miscible  
13      organic solvent is ethyl alcohol.

12        2.      The liquid composition of Claim 1 wherein the water miscible  
13      organic solvent is ethyl alcohol.

3. The liquid composition of Claim 2 that contains approximately

15 15 to 60 % ethyl alcohol.

16 4. The liquid composition of Claim 3 that contains approximately  
17 50 % ethyl alcohol.

18        5.    The liquid composition of Claim 1 that contains approximately 2  
19    to 10 % polyethylene glycol.

20        6.      The liquid composition of Claim 5 that contains approximately 5  
21      % polyethylene glycol

22        7.      The liquid composition of Claim 5 wherein the polyethylene  
23      glycol has a molecular weight of approximately 8,000 D.

24           8.     The liquid composition of Claim 1 that contains approximately  
25   0.1 to 0.7 % heparin.

26 9. The liquid composition of Claim 8 that contains approximately  
27 0.5% heparin.

28 10. An aqueous liquid composition adapted for treating autologous  
29 biological tissue for modifying its tissue reactivity and for rendering the tissue

1 temporarily more rigid than in its natural state, the composition comprising:  
2 approximately 15 to 60 % by volume of ethyl alcohol;  
3 approximately 2 to 10 % by weight of polyethylene glycol of a  
4 molecular weight of approximately 8,000 D;  
5 approximately 0.1 to 0.7 % by weight of heparine, and  
6 the balance of the composition substantially consisting of water.

7 11. The liquid composition of Claim 10 comprising approximately  
8 50 % ethyl alcohol, approximately 5 % polyethylene glycol and approximately  
9 0.5 % heparine.

10 12. A method for modifying the tissue reactivity of an autologous  
11 tissue freshly obtained from a host mammal and for rendering the tissue  
12 temporarily more rigid than in its native state, the method comprising:  
13 exposing said autologous tissue to a liquid composition, comprising:  
14 approximately 10 to 70 % by volume of a water miscible non-toxic  
15 organic solvent selected from the group consisting of an alcohol having 1 to 3  
16 carbons, acetone, acetonitrile and methyl ethyl ketone;  
17 approximately 2 to 30 % by weight of polyethylene glycol having a  
18 molecular weight in the range of approximately 6,000 to 15,000 D;  
19 approximately 0.01 to 1.0 % by weight of heparin, and  
20 the balance of the composition substantially consisting of water.

21 13. The method of Claim 12 where in the liquid composition the  
22 water miscible organic solvent is ethyl alcohol.

23 14. The method of Claim 13 where the liquid composition contains  
24 approximately 15 to 60 % ethyl alcohol.

25 15. The method of Claim 13 where the liquid composition contains  
26 approximately 2 to 10 % polyethylene glycol.

27 16. The method of Claim 13 where the liquid composition contains  
28 approximately 0.1 to 0.7 % heparin.

29 17. The method of Claim 13 where the liquid composition contains

1 approximately 50 % ethyl alcohol, approximately 5 % polyethylene glycol of a  
2 molecular weight of approximately 8,000 D and approximately 0.5 % heparin.

3       **18.**    The method of Claim 12 where the autologous tissue freshly  
4 obtained from a host mammal comprises a biological membrane.

5       **19.**    The method of Claim 12 where the autologous tissue is selected  
6 from a group consisting of peritoneum, pericardium, pleura and tendon.

7       **20.**    The method of Claim 12 wherein the step of exposing the  
8 autologous tissue to the liquid composition is by immersing the tissue in the  
9 liquid composition.

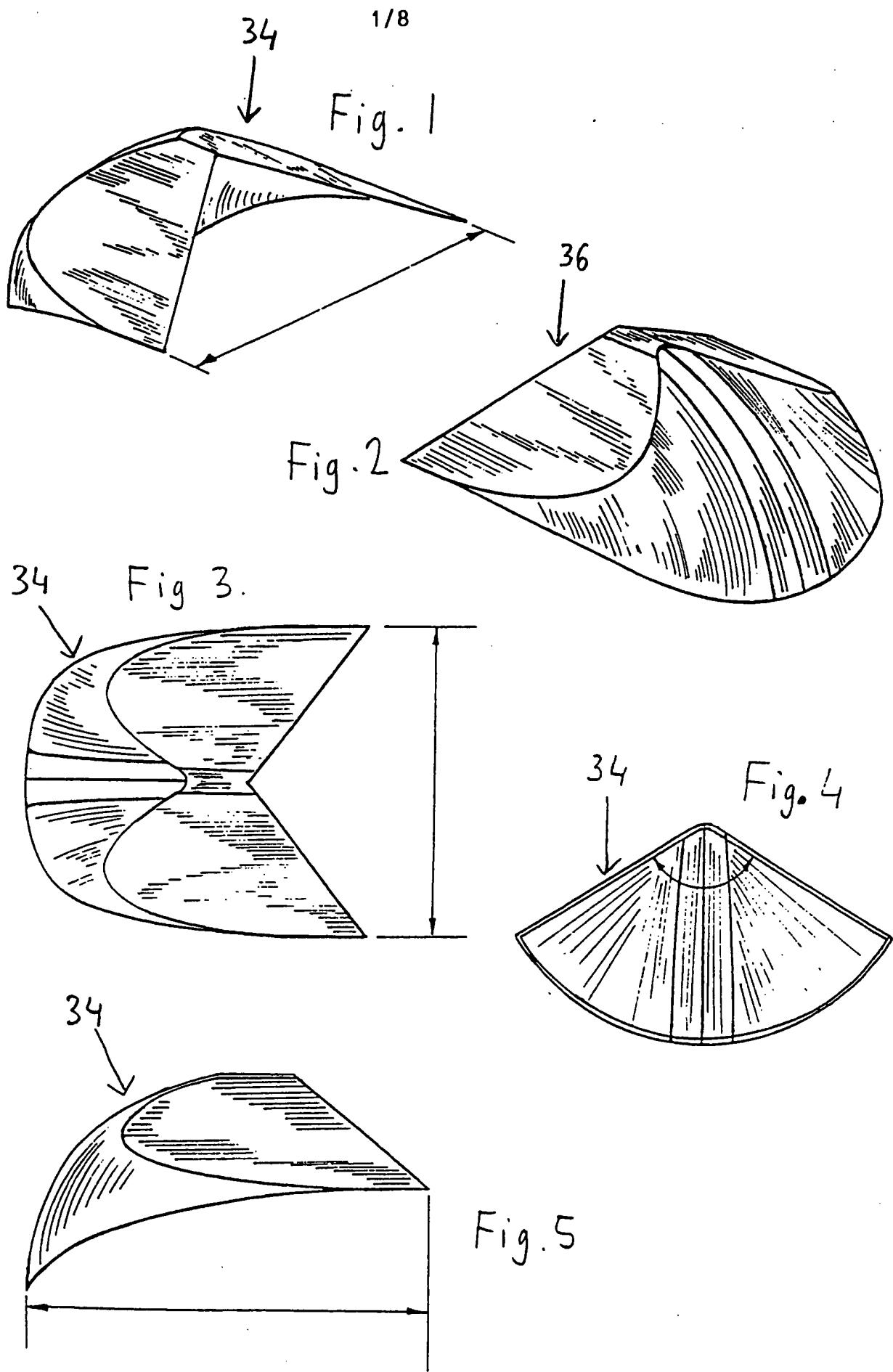
10       **21.**    The method of Claim 20 further comprising the step of placing  
11 the autologous tissue in a mold.

12       **22.**    The method of Claim 21 wherein the autologous tissue is placed  
13 in the mold before the tissue is exposed to the liquid composition, and wherein  
14 the tissue is exposed to the liquid composition while it is held in said mold,  
15 thereby forming said tissue into a predetermined configuration.

16       **23.**    The method of Claim 22 further comprising the step of trimming  
17 excess autologous tissue by cutting while said tissue is still in the mold and  
18 after it has been exposed to said liquid composition for at least approximately  
19 2 to 8 minutes.

20       **24.**    The method of Claim 23 further comprising the step of  
21 implanting the trimmed tissue into the host.

22       **25.**    The method of Claim 24 further comprising the step of irrigating  
23 the implanted tissue with isotonic saline solution thereby causing the physical  
24 properties of the tissue to return to their substantially native state.



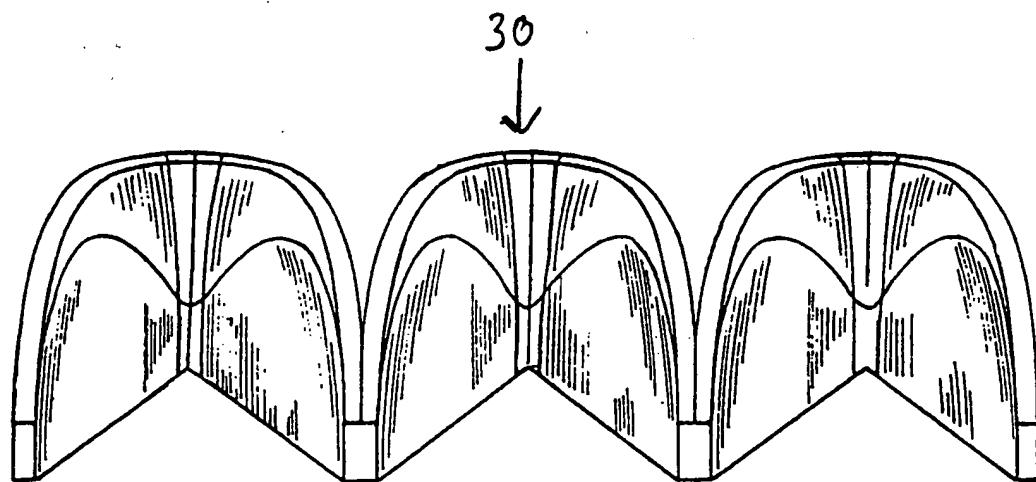


Fig. 6

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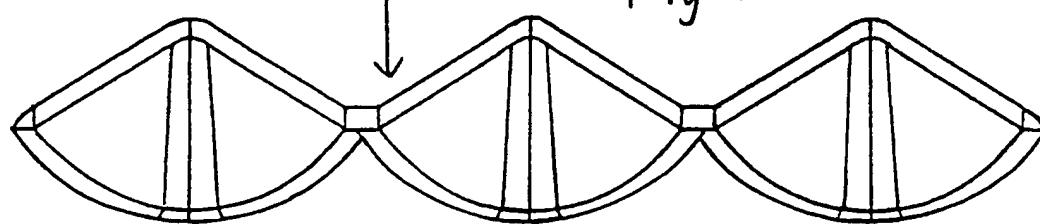
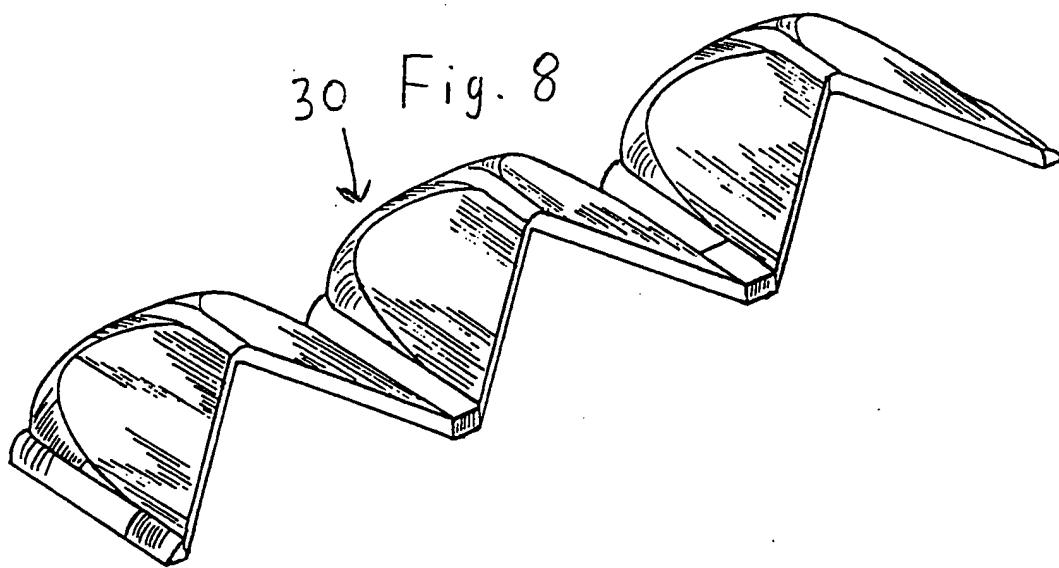


Fig. 7.

30 Fig. 8



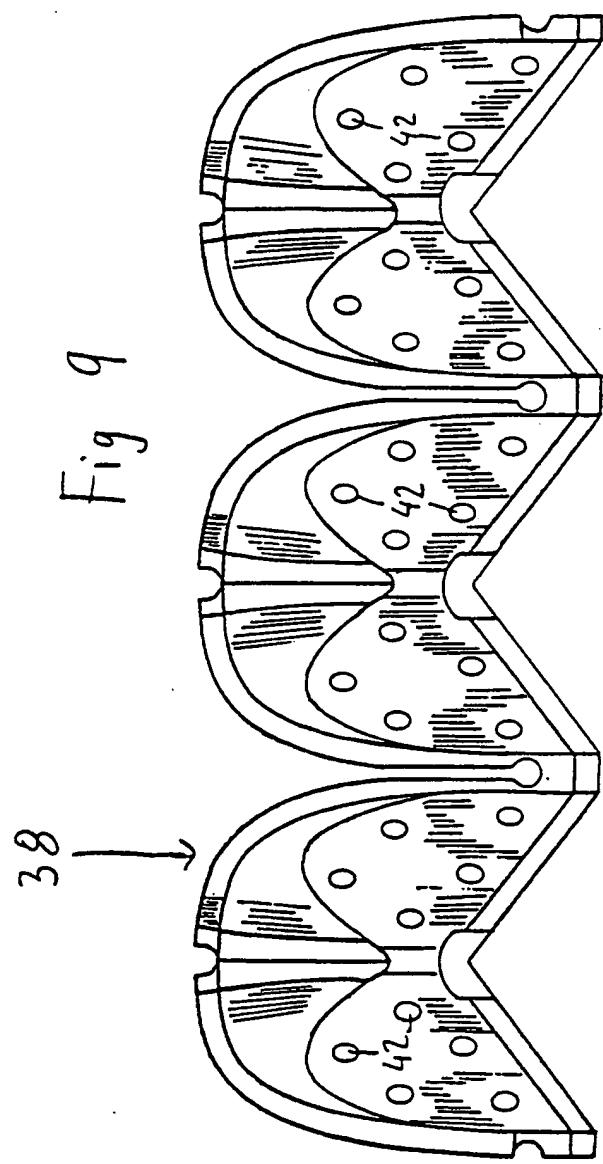


Fig. 9

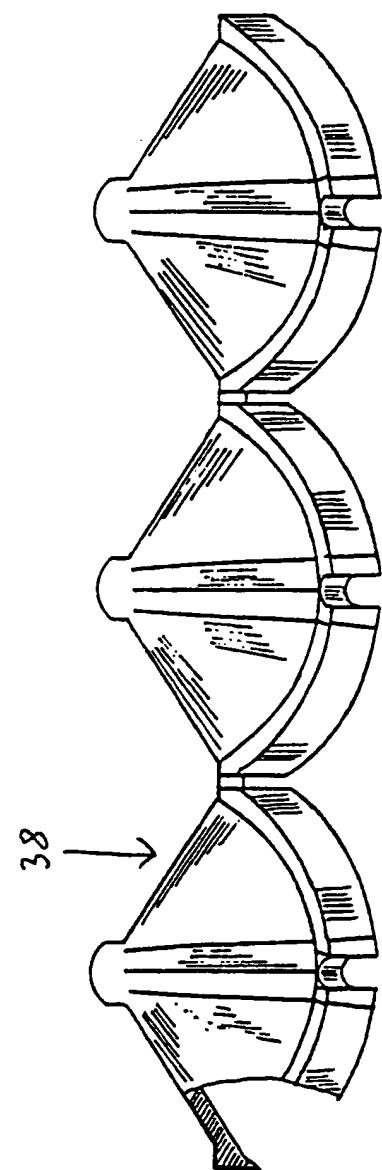


Fig. 11

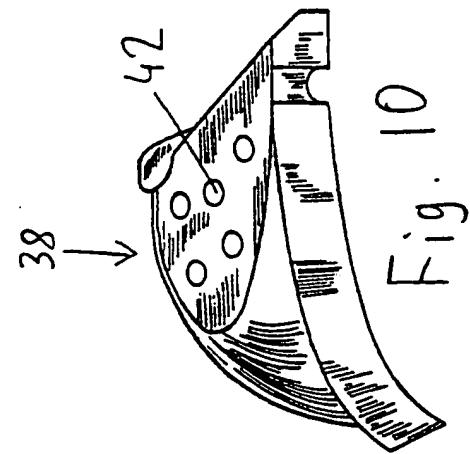


Fig. 10

Fig. 12

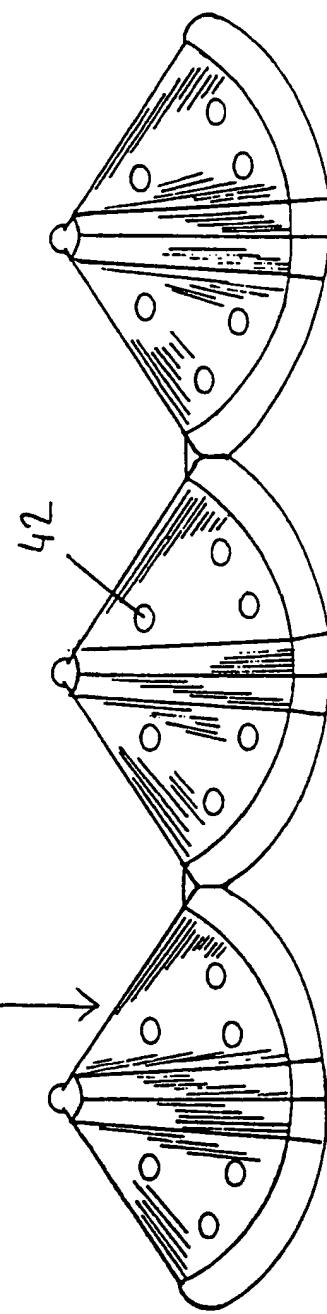
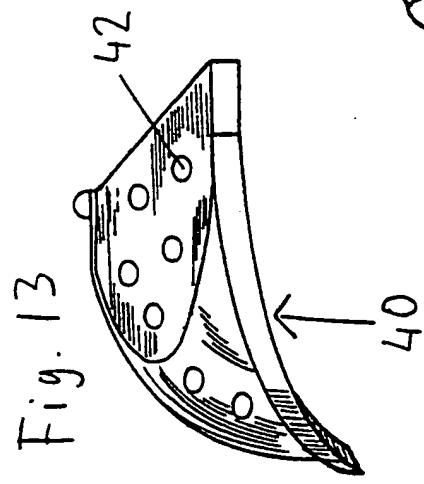
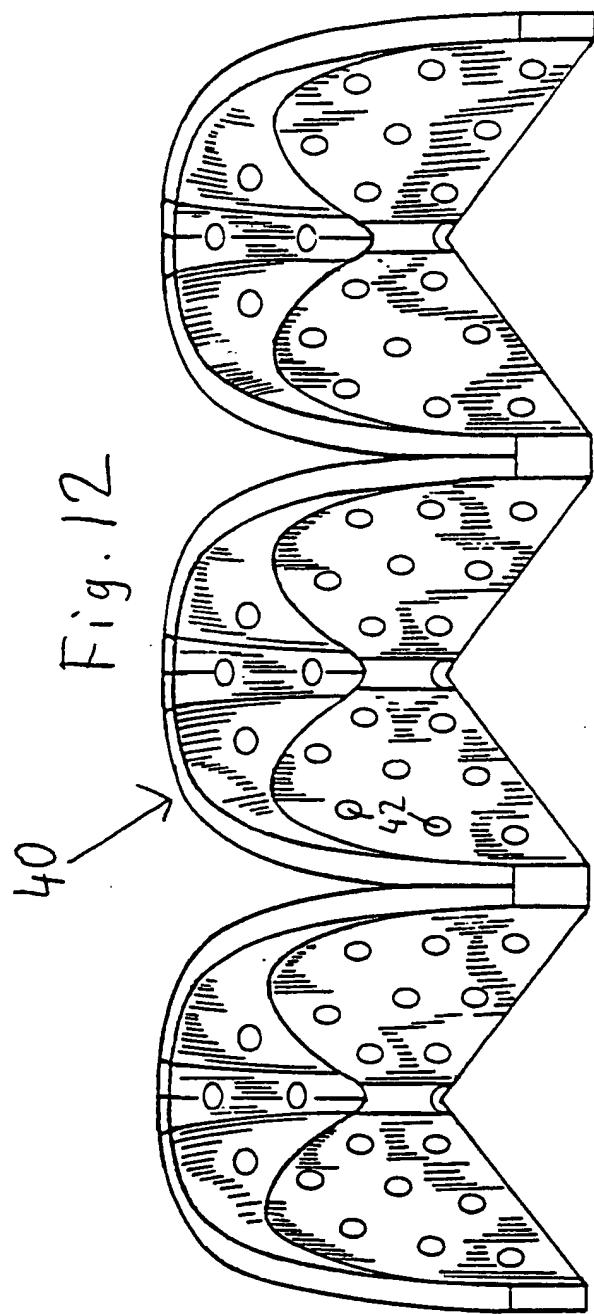


Fig. 14

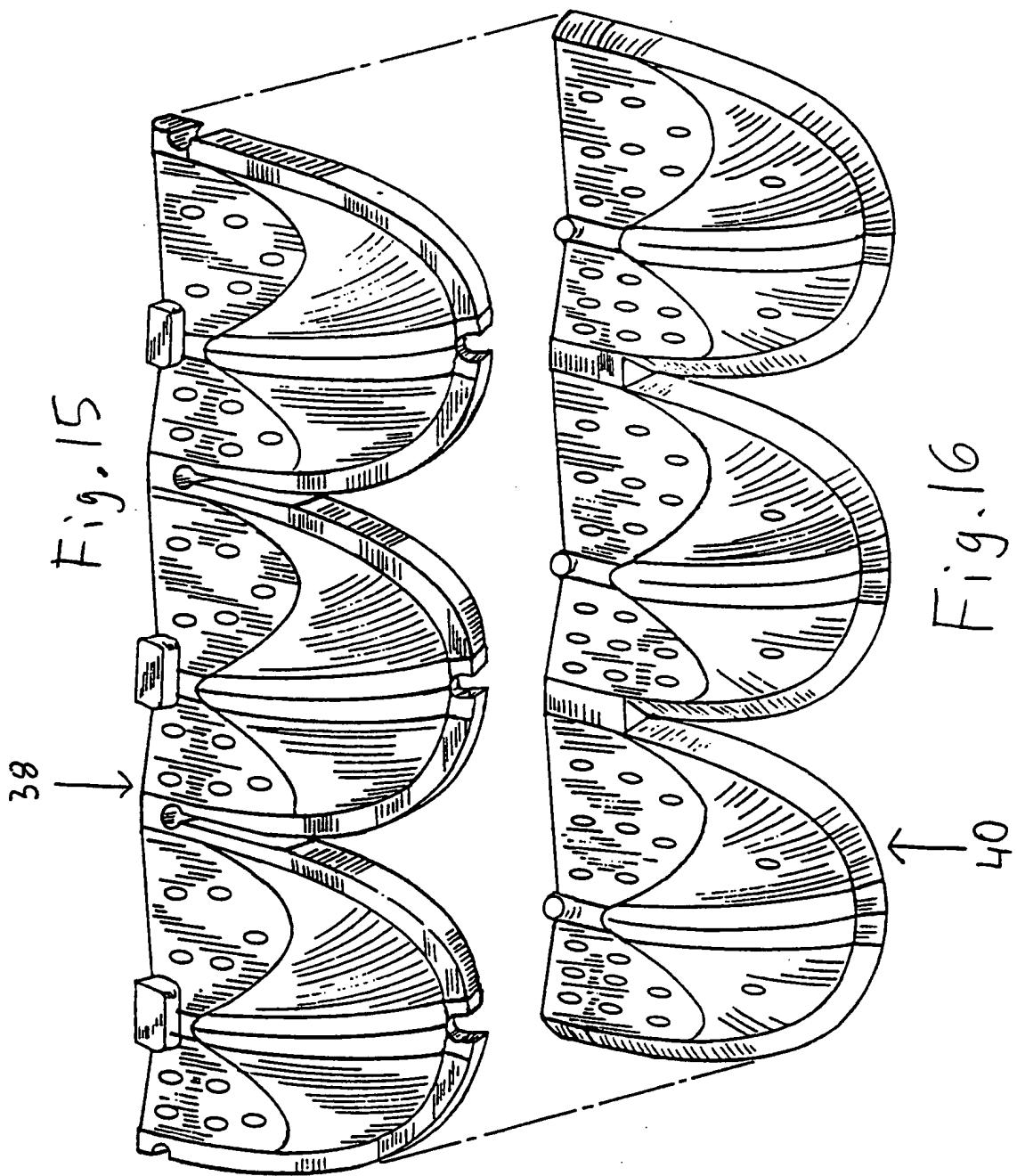
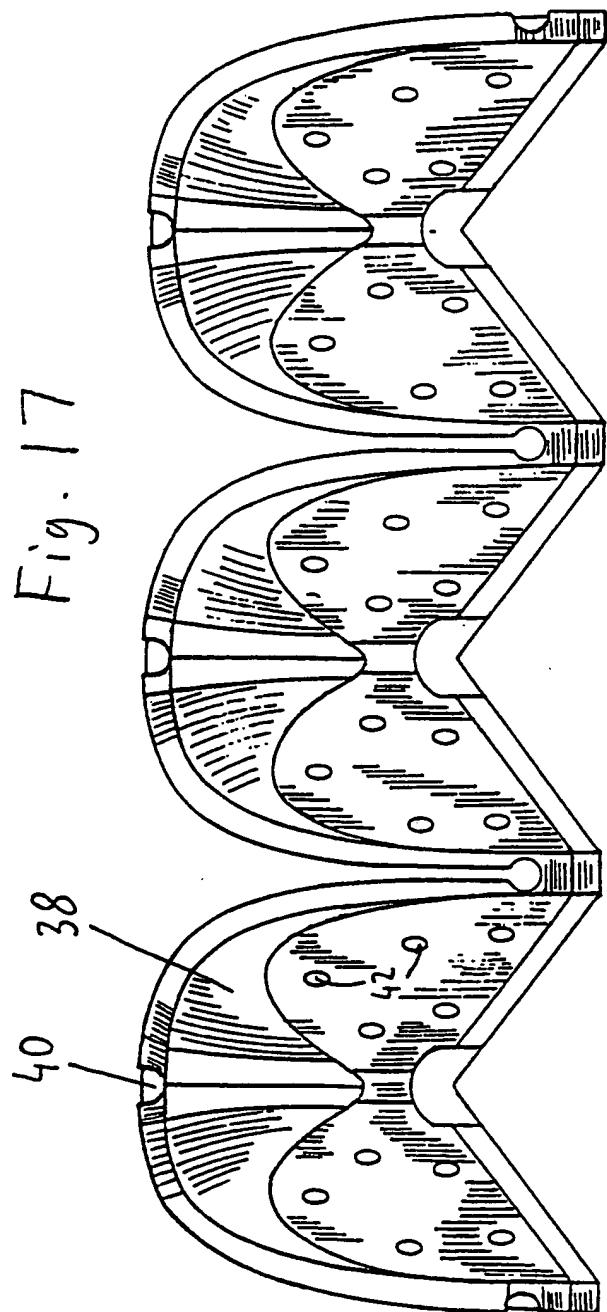
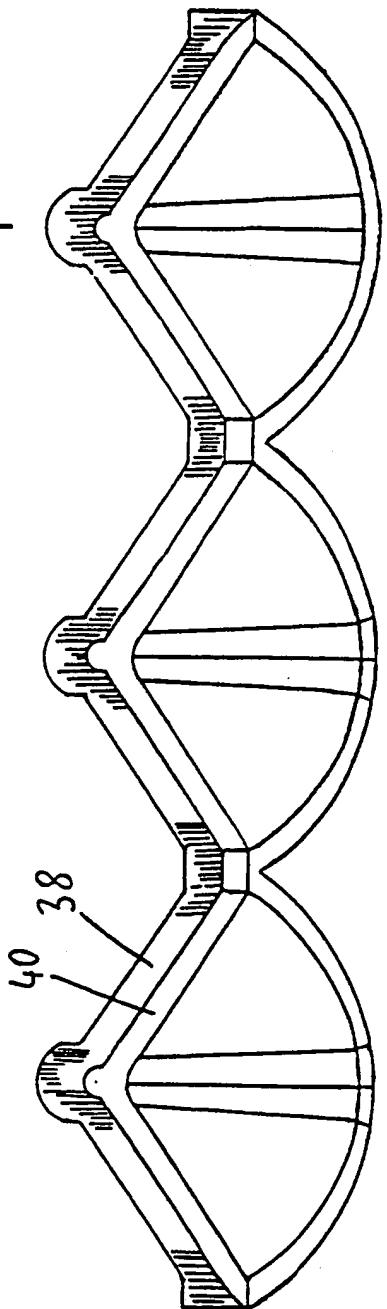


Fig. 17

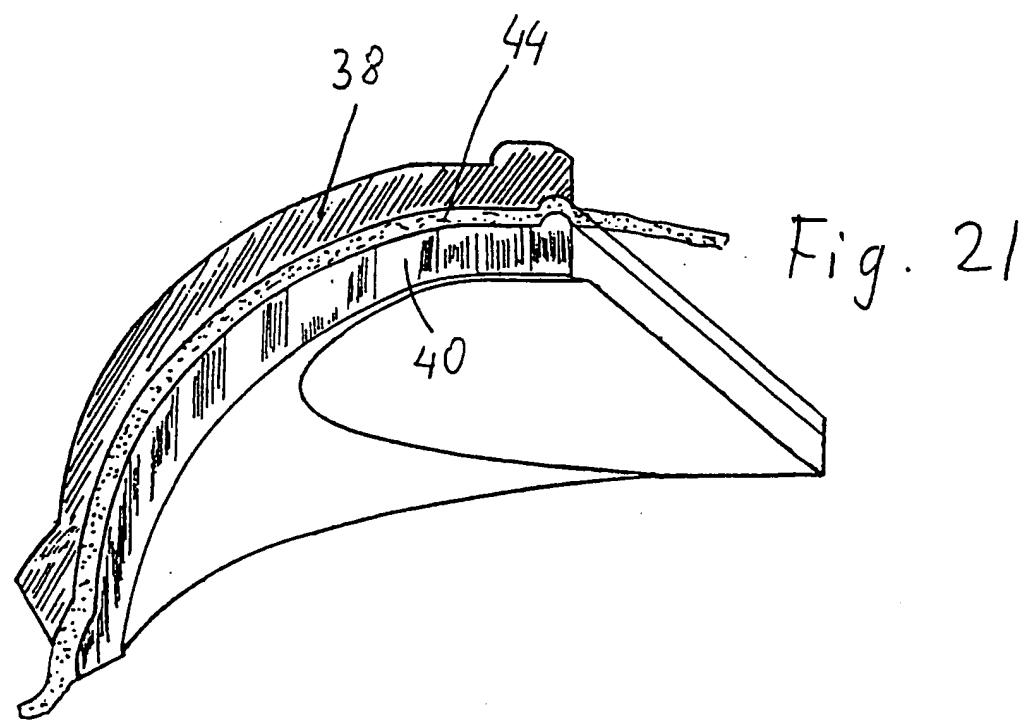
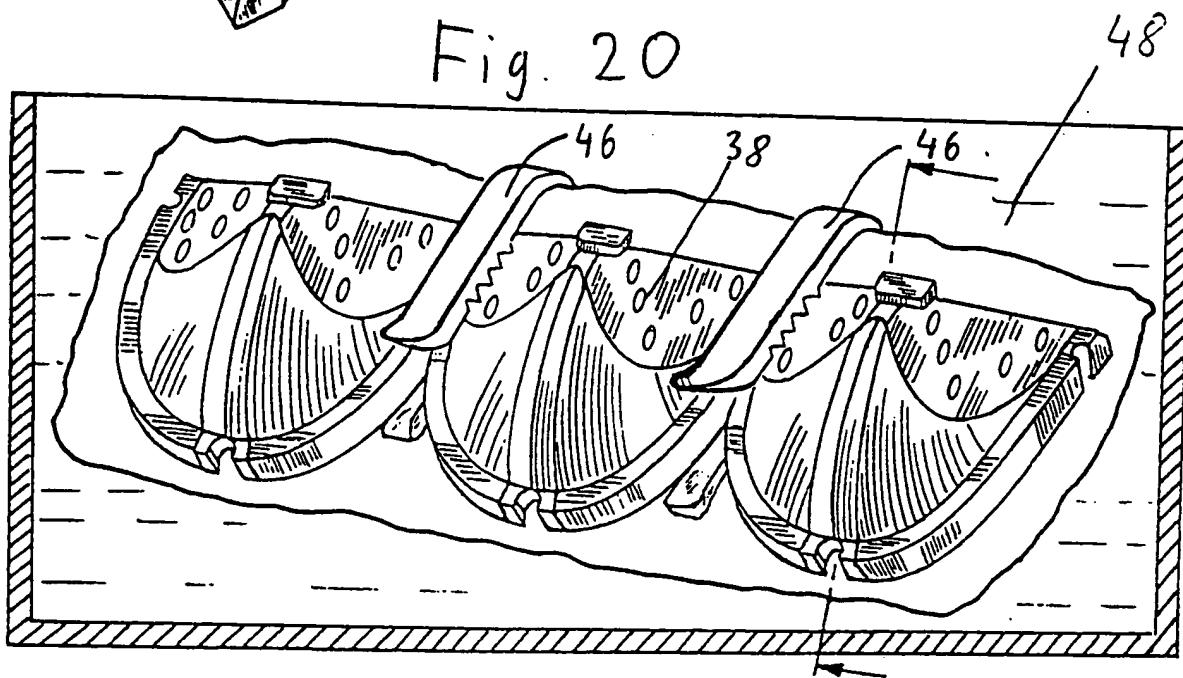
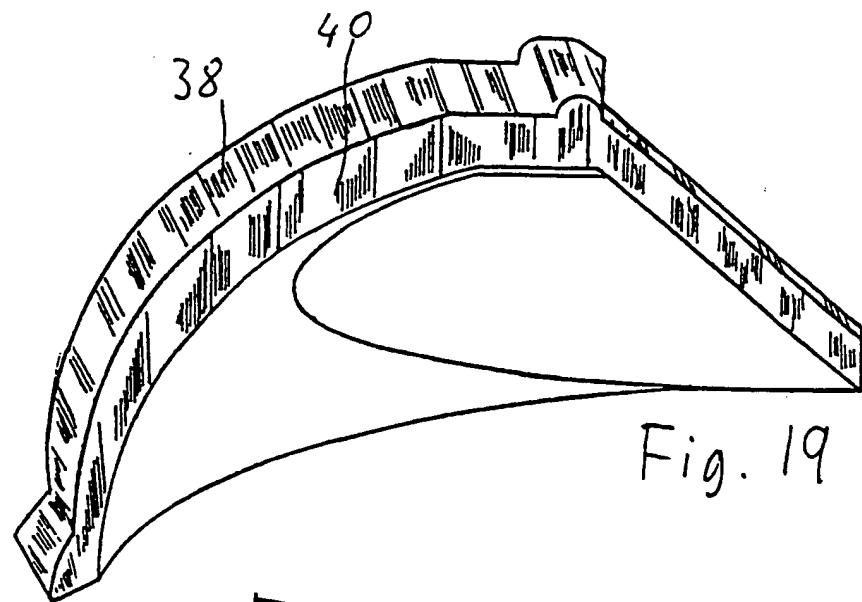


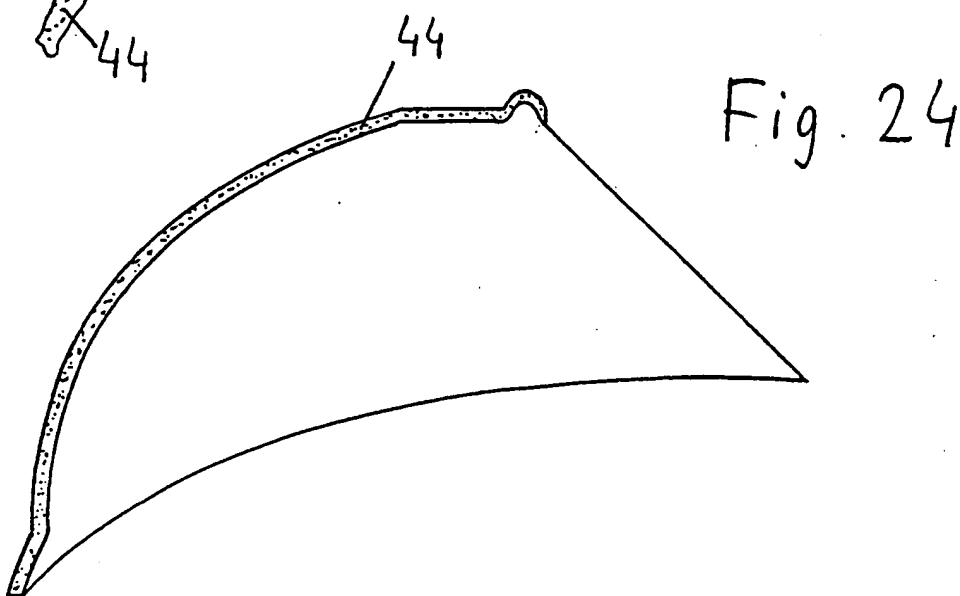
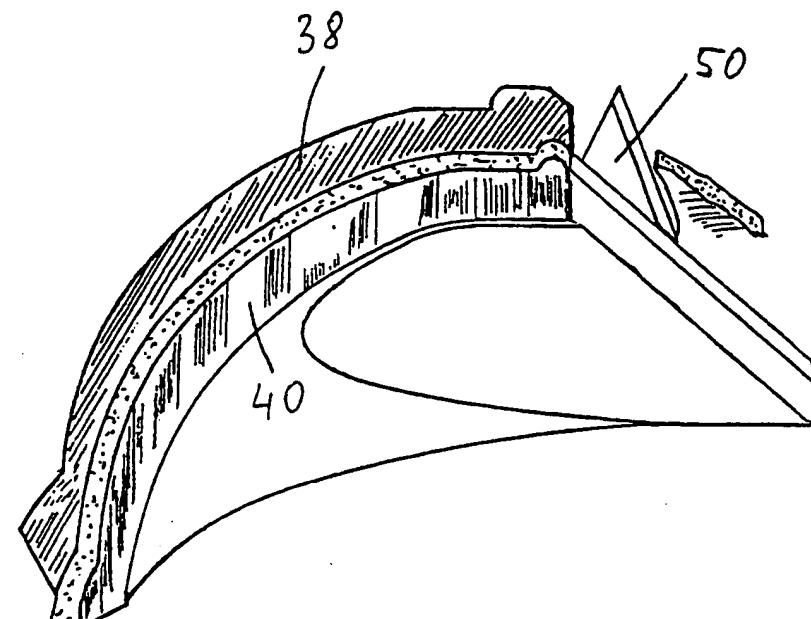
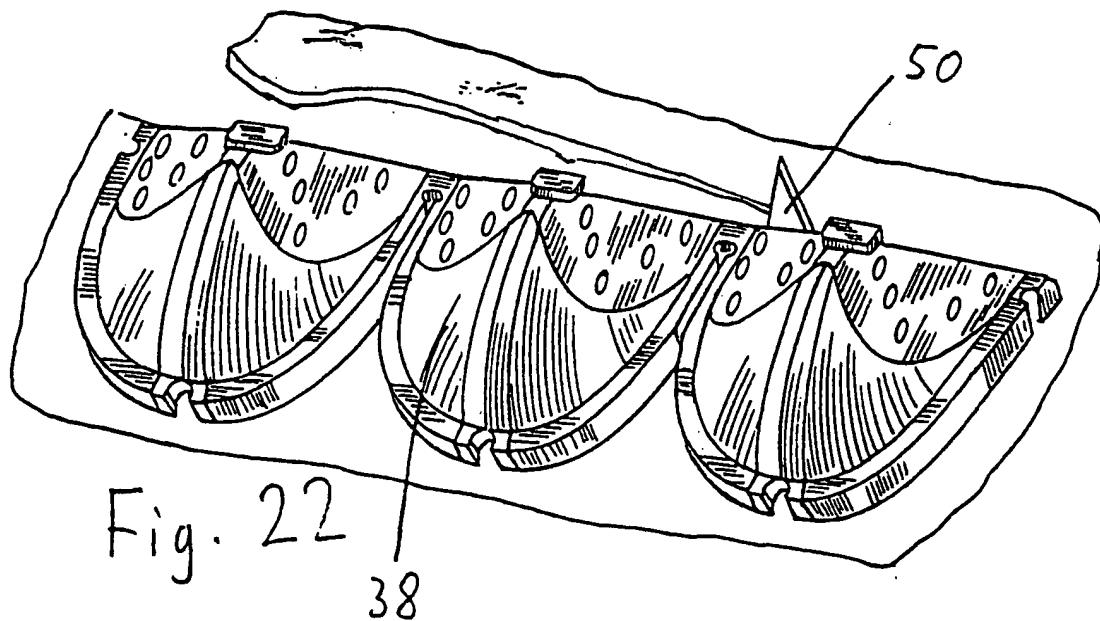
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Fig. 18





## INTERNATIONAL SEARCH REPORT

Int. Application No  
PCT/US 00/41142

A. CLASSIFICATION OF SUBJECT MATTER  
IPC 7 A61L27/36 A01N1/02 A61F2/24

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)  
IPC 7 A61L A61F A01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, PAJ

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P, X	WO 99 66967 A (INT HEART INST OF MONTANA FOUN) 29 December 1999 (1999-12-29) the whole document ----	1-25
A	WO 98 07452 A (SULZER VASCUTEK LTD ;WALKER DONALD FRANCIS (GB)) 26 February 1998 (1998-02-26) claims; examples ----	1-25
A	US 5 558 875 A (WANG SU) 24 September 1996 (1996-09-24) cited in the application claims ----	1-25
A	WO 97 32472 A (BARBARASH LEONID SEMENOVICH ;ZHURAVLEVA IRINA JURIEVNA (RU); GANTI) 12 September 1997 (1997-09-12) abstract ----	1-25



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Patent family members are listed in annex.

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Date of the actual completion of the international search

1 March 2001

Date of mailing of the international search report

08/03/2001

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ESPINOSA, M

**INTERNATIONAL SEARCH REPORT****Information on patent family members**I. National Application No  
**PCT/US 00/41142**

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
WO 9966967	A 29-12-1999	AU 4829899	A	10-01-2000
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